

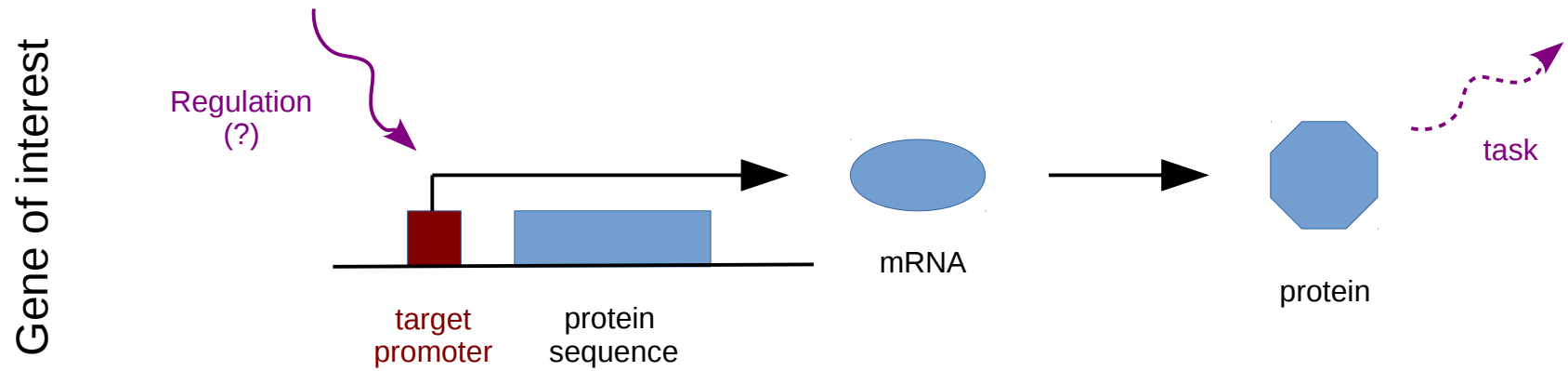


# Extrinsic noise estimation in single-cell gene expression

Eugenio Cinquemani  
*INRIA Grenoble – Rhône-Alpes*

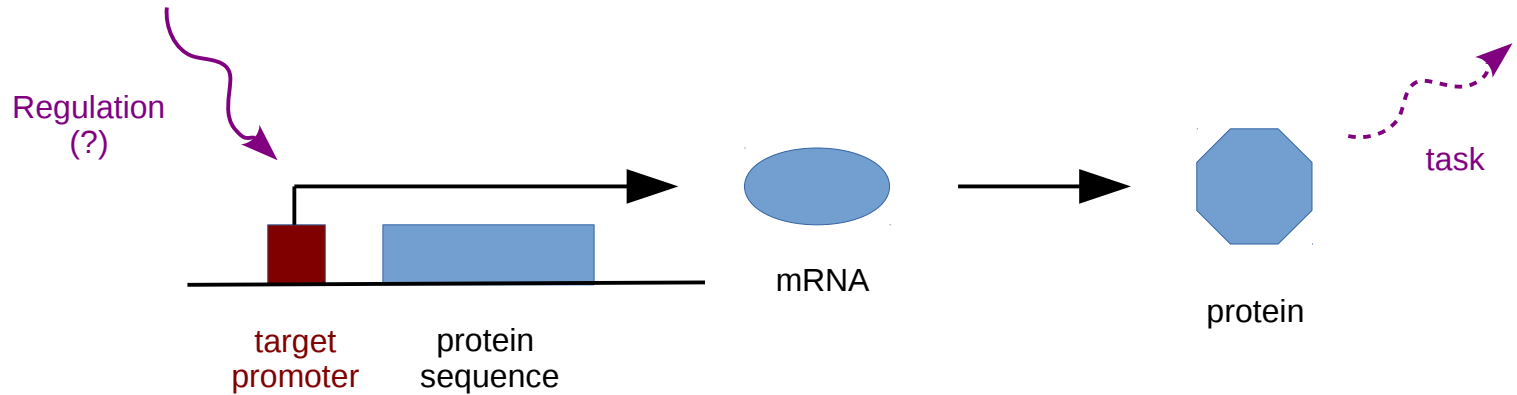
ASSTABIO  
Rennes, March 23, 2018

# Gene expression

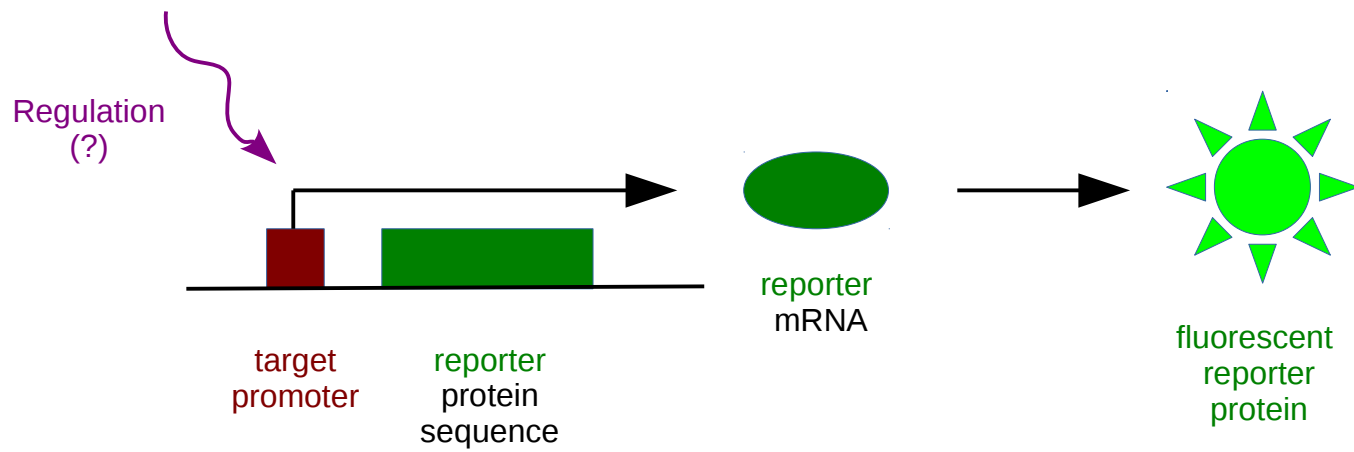


# Gene expression and fluorescence reporting

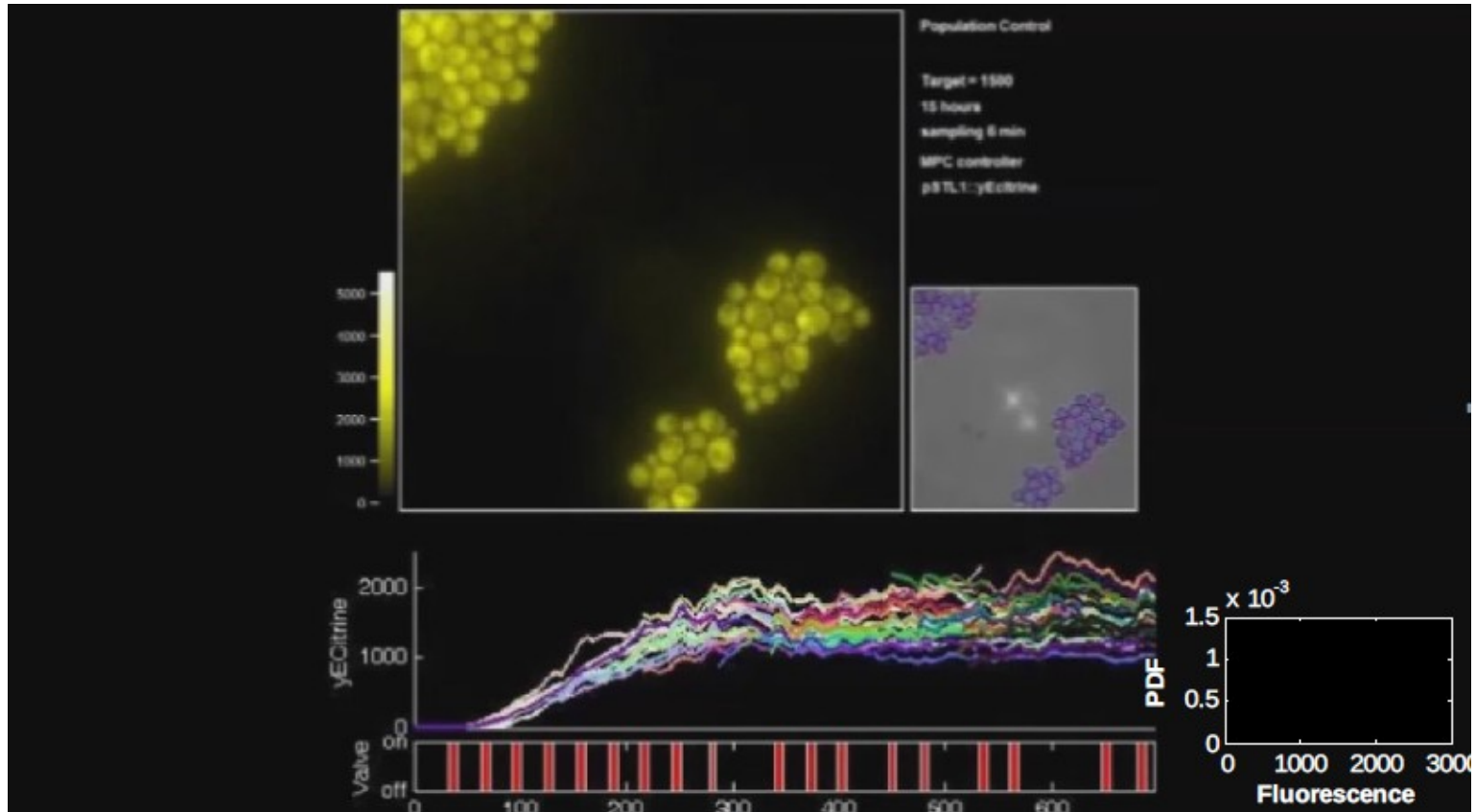
Gene of interest



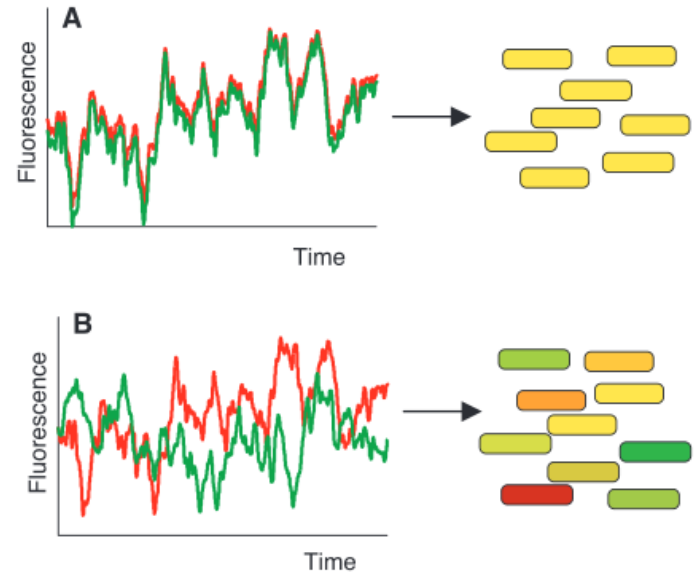
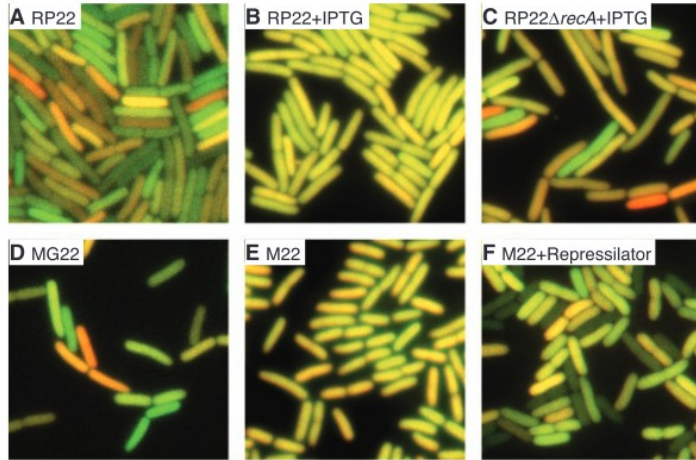
Gene reporter system



# Gene expression variability



# Intrinsic vs. extrinsic noise



(Elowitz et al, Science 2002)

- Intrinsic : Random transcription and translation events
- Extrinsic : Other sources of variability  
(parameters, promoter activity, ... )

# Outline

- Cell-to-cell parameter variability in osmotic shock response  
(Llamosi et al., PloS 2016)
- Stochastic variability in promoter activity  
(Cinquemani, arXiv 2017 and under review)

RESEARCH ARTICLE

# What Population Reveals about Individual Cell Identity: Single-Cell Parameter Estimation of Models of Gene Expression in Yeast

Artémis Llamasi<sup>1,2</sup>, Andres M. Gonzalez-Vargas<sup>3</sup>, Cristian Versari<sup>4</sup>,  
Eugenio Cinquemani<sup>5</sup>, Giancarlo Ferrari-Trecate<sup>3†\*</sup>, Pascal Hersen<sup>2†\*</sup>, Gregory Batt<sup>1†\*</sup>



1 Inria Saclay - Ile-de-France, Palaiseau, France, 2 Laboratoire Matière et Systèmes Complexes, UMR 7057, Université Paris Diderot & CNRS, Paris, France, 3 Dipartimento di Ingegneria Industriale e dell'Informazione, Università degli Studi di Pavia, Pavia, Italy, 4 Laboratoire d'Informatique Fondamentale de Lille, UMR 8022, Université de Lille 1 & CNRS, Villeneuve d'Ascq Cedex, France, 5 INRIA Grenoble - Rhône-Alpes, Montbonnot, France

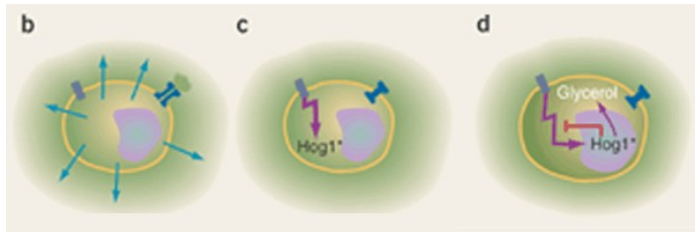
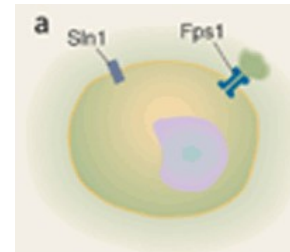
## Intercellular variability in osmotic shock response

*inria*  
informatics mathematics

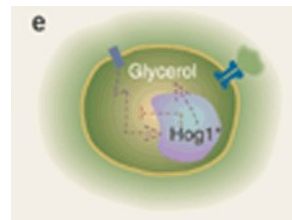
# Target system :

## Osmotic shock response in yeast

- Low-osmolarity environment :  
Cells are fine
- Increase of environmental osmolarity :  
Cells feel bad and react



- Eventually, cell adapt to the new medium



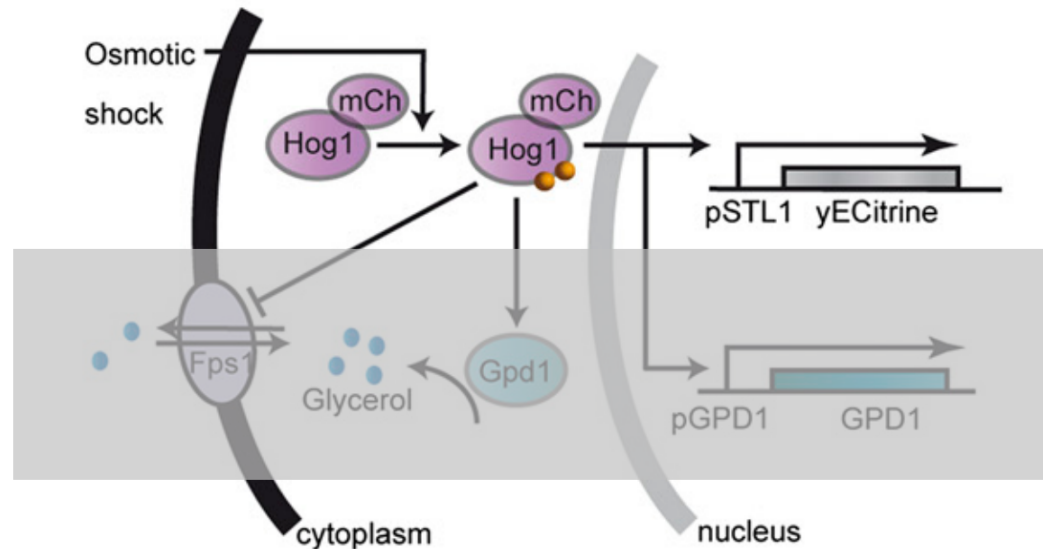
(b) Loss of turgor, water rushes out of the cells.  
(c) glycerol efflux channel closed, accumulation of glycerol, Hog1 phosphorylation.  
(d) Hog1 migrates to nucleus, upregulates expression in several genes

(e) cell re-inflation decreases HOG signaling. Phosphatases deactivate HOG pathway, turgor is restored and Fps1 reopens.



# HOG1 pathway : Gene expression monitoring

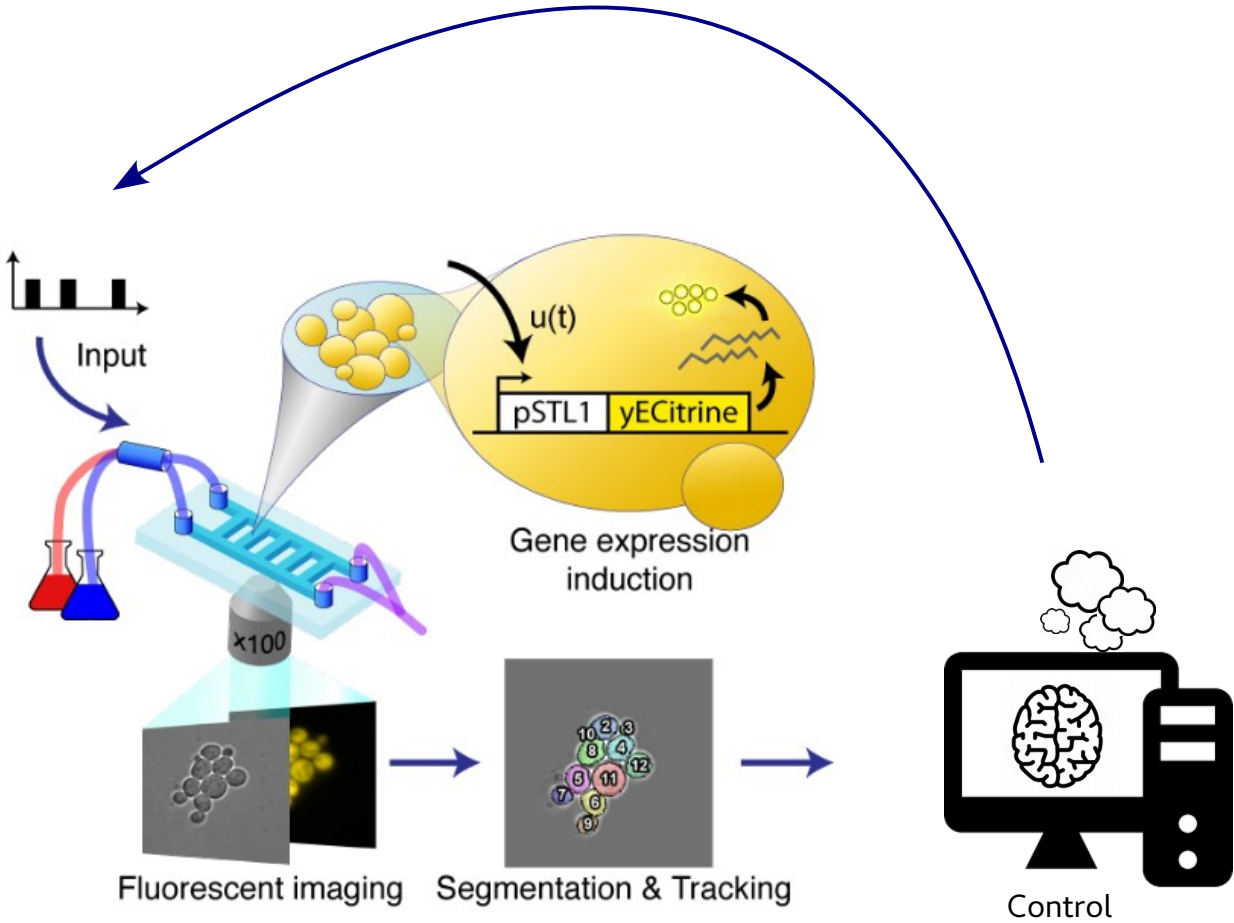
- Focus on expression dynamics of osmosensitive genes
- Fluorescent reporter monitoring of STL1 activity under short osmolarity pulses :



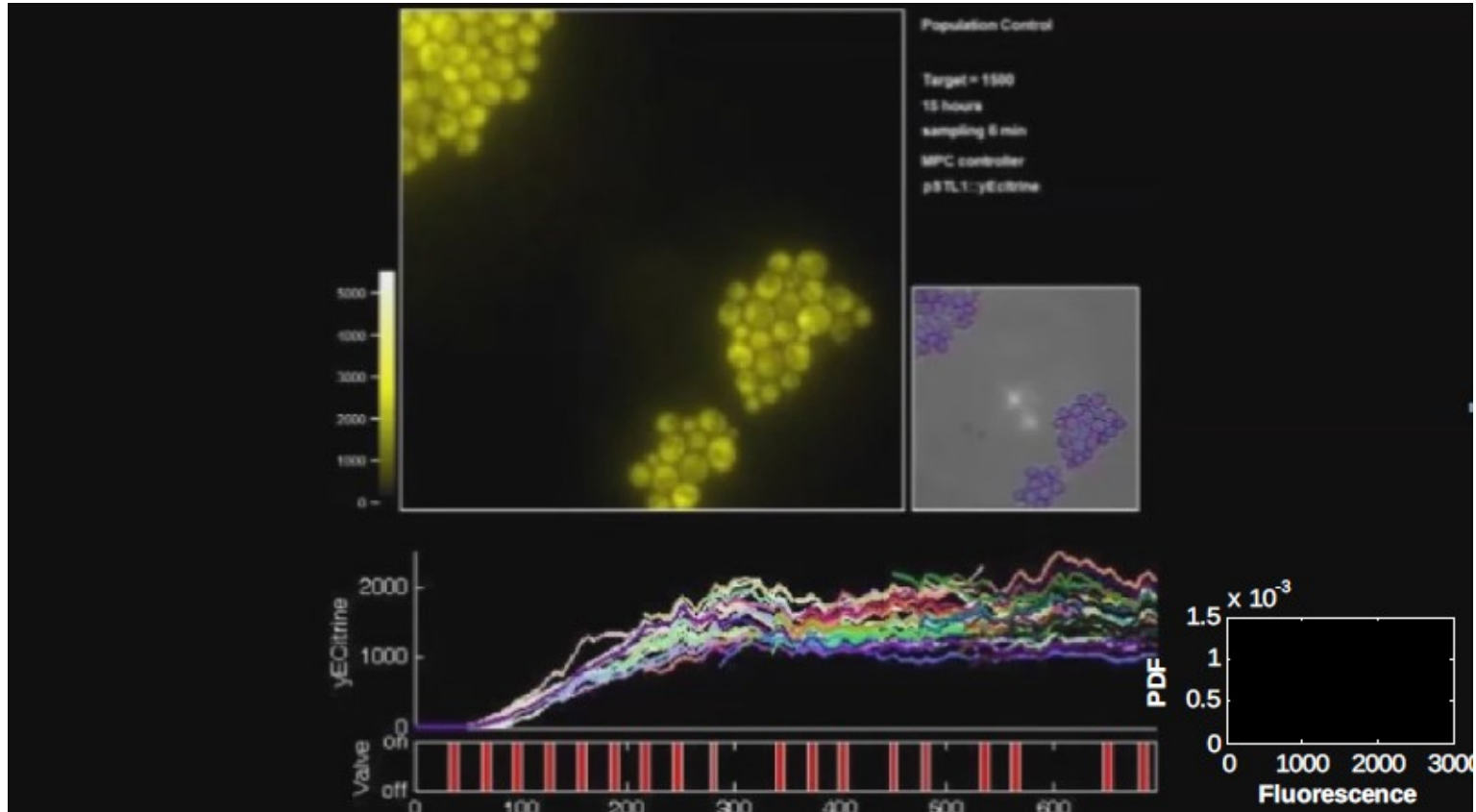
(Uhlendorf et al., *PNAS* 2012)

- Feedback regulation is cut or not triggered (adaptation prevented)

# Experimental setup



# Life according to single cells...



- Question #1 : How to better control this ?
- Question #0: How to make sense of (i.e. model) this ?

# Single-cell kinetic model

Input: Osmotic shock signal  $u(t)$

Gene expression:

$$\begin{aligned}\dot{m}(t) &= k_m u(t) - g_m m(t), \\ \dot{p}(t) &= k_p m(t) - g_p p(t),\end{aligned}$$

Protein maturation:

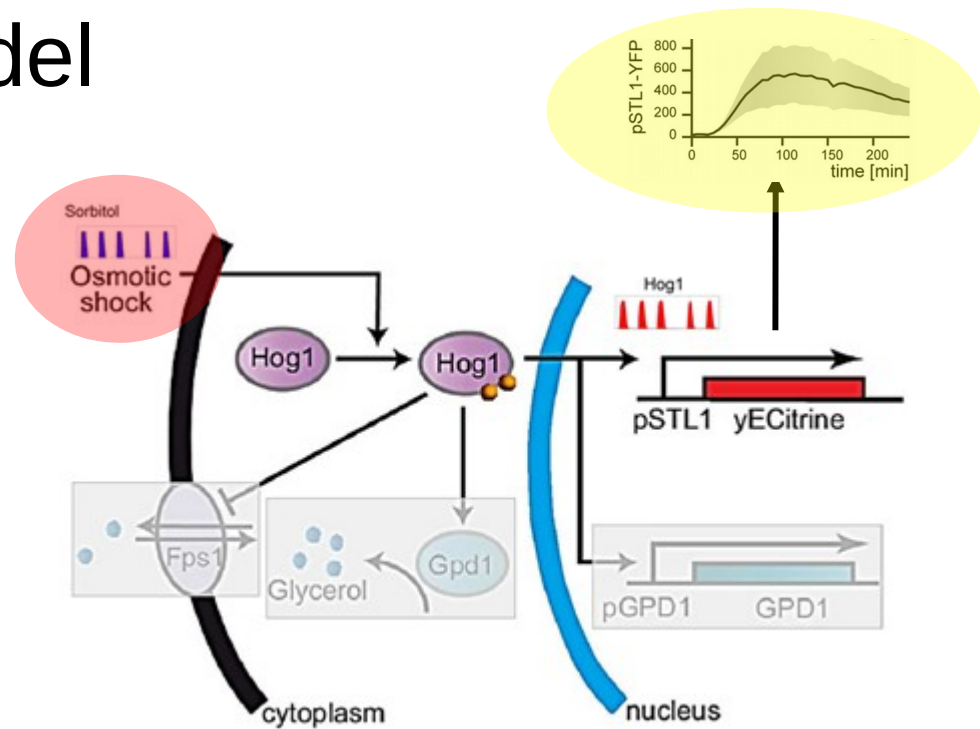
$$f(t) = e^{-g_p \tau} p(t - \tau).$$

Measurement noise intensity:

$$h(t) = (\varepsilon_a + f(t)\varepsilon_b)$$

Output:

$$y(t) = f(t) + h(t)\eta$$



Parameters:

$$\theta = (k_p, g_p, k_m, g_m), \tau, \text{ initial conditions}$$

Our focus:

$$k_{mp} = k_m \cdot k_p, g_m, g_p$$

# Mixed-Effects paradigm

- $N$  cells are **different individuals** of the **same population**
- Different **cell parameters** following a **common population distribution**

$$\theta_1, \dots, \theta_N \text{ i.i.d.}, \quad \log \theta_i \sim \mathcal{N}(\mu, \Sigma)$$

- *Inference from data* : Learn **population statistics** from the ensemble of the **individual-cell data**

$$Y = \{y_i(t) : \forall t, i = 1, \dots, N\}$$

- *Key point* : Every individual contributes to learning population features, and so it helps learning features of all other individuals

# Mixed-Effects model inference

- Estimate population statistics by marginal likelihood maximization

$$(\hat{\mu}, \hat{\Sigma}) \leftarrow \max_{\mu, \Sigma} p(y|\mu, \Sigma), \quad p(y|\mu, \Sigma) = \prod_i \int p(y_i|\theta_i) p(\theta_i|\mu, \Sigma) d\theta_i$$

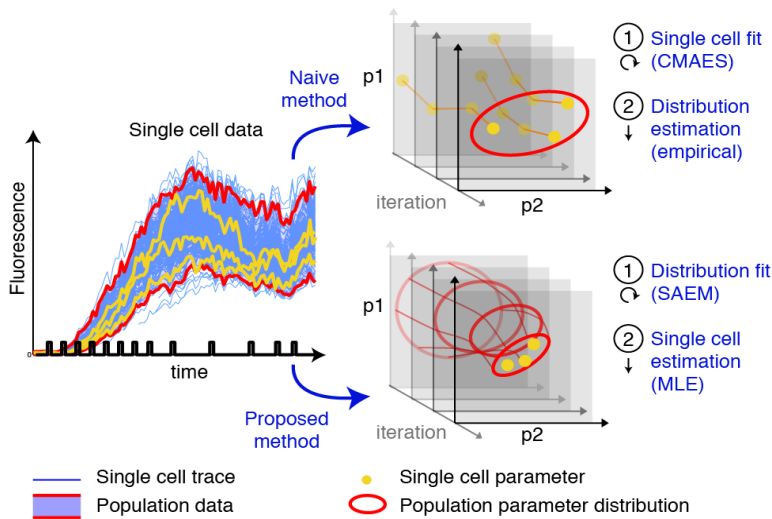
- Estimate single-cell parameters via Maximum-A-Posteriori

$$\hat{\theta}_i \leftarrow \max_{\theta_i} p(\theta_i|y_i, \hat{\mu}, \hat{\Sigma}), \quad p(\theta_i|y_i, \hat{\mu}, \hat{\Sigma}) \propto p(y_i|\theta_i) \cdot p(\theta_i|\hat{\mu}, \hat{\Sigma})$$

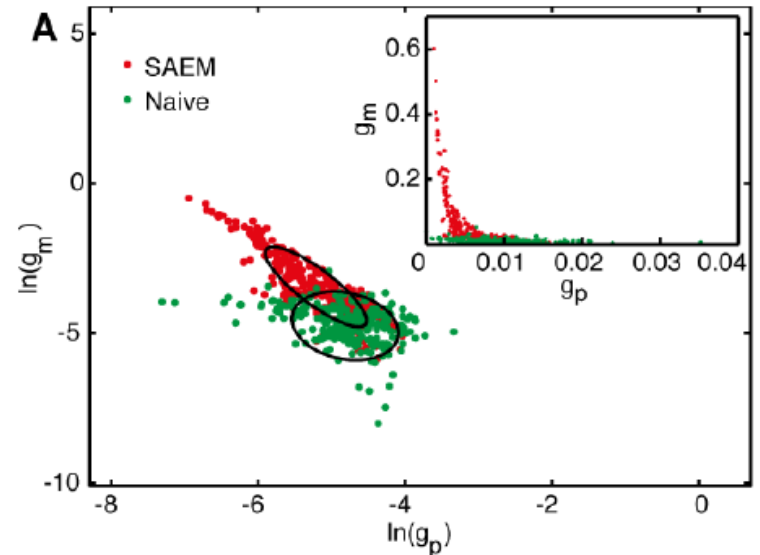
- Effective implementation (SAEM):  
Stochastic Approximation of Expectation-Maximization

# Mixed-Effects vs. naive inference

- Naive approach: Empirical parameter statistics from individual cell fits



- Parameter distribution from ME inference more structured and compact



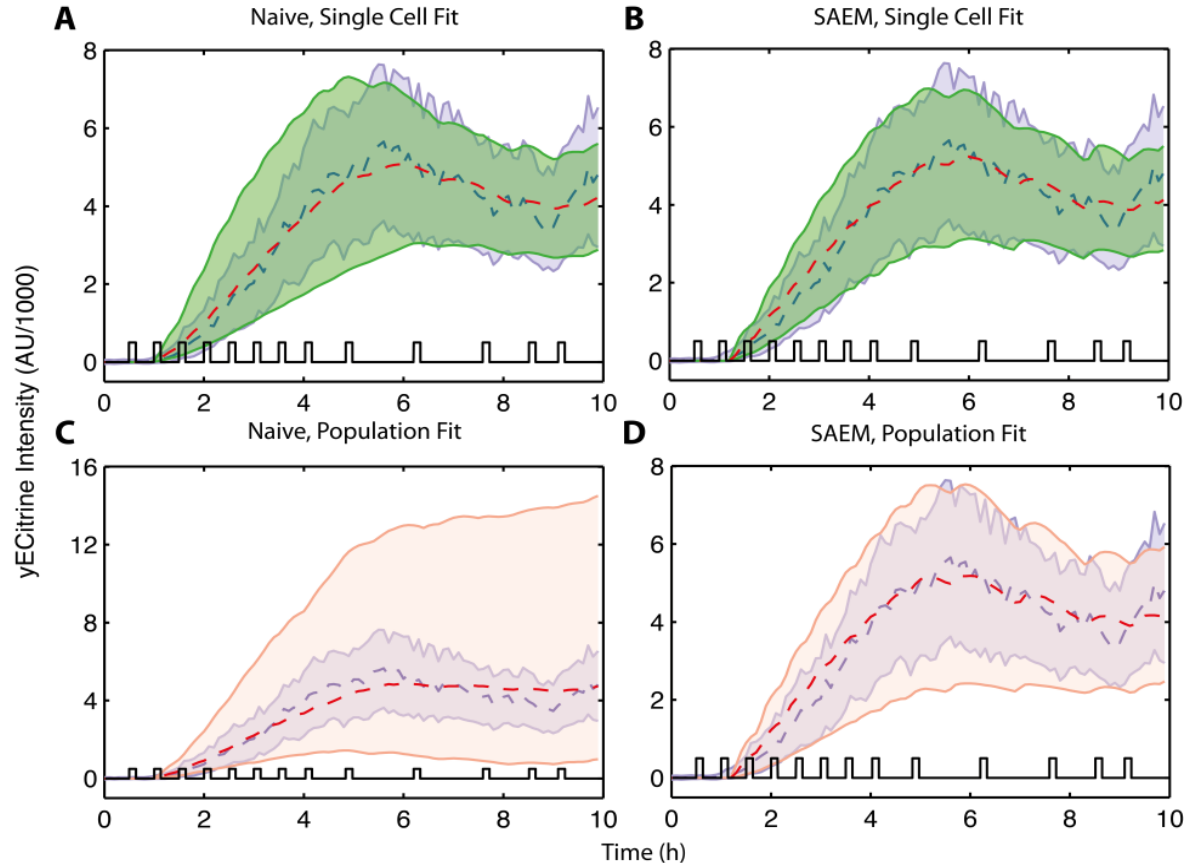
**B**

	Parameter distribution structure	Parameter distribution spread
SAEM	0.62	6.7
Naive	0.19	37

# ME inference *does* retrieve population statistics

Single-cell trajectories from parameter estimates (confidence bounds)

Resimulation of single-cell trajectories via parameter resampling (confidence bounds)



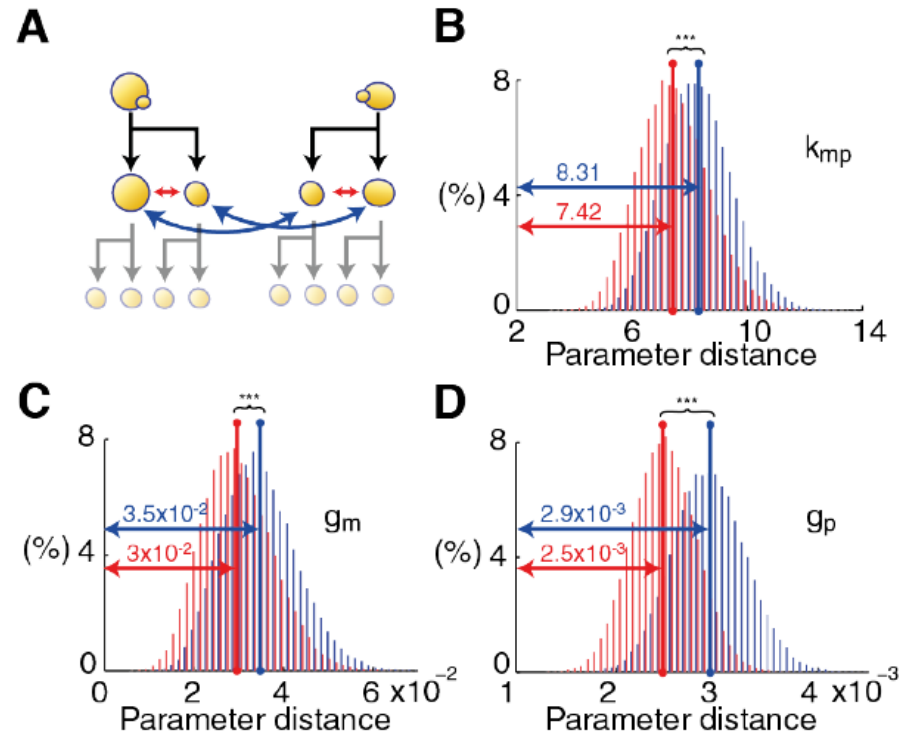
Naive approach

ME approach



# Mother-daughter parameter inheritance

- Daughter cell parameters statistically closer to those of their mothers
- A formal yet 'irrefutable' observation to be investigated further



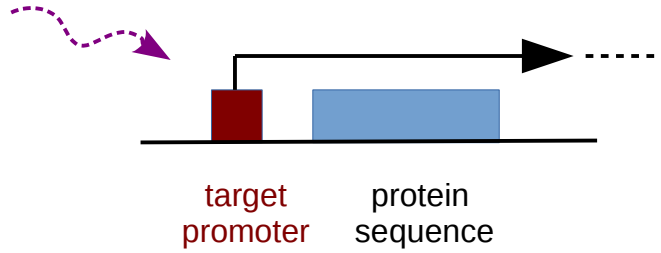
**Fig. 7: Parameter values of individual cells are statistically closer to the parameters of their own mother than to the parameters of another mother cell.** (A) The distance between parameters of related mother and daughter cells (MD) and non-related mother and daughter cells (nMD) were compared. (B-D) Distribution for each parameter of the average distance between 40 pairs of MD (red) and nMD (blue) for 50000 combinations obtained by bootstrapping (Text S1). All parameters are closer between mothers and daughters than on average (\*\*\*)  $p$ -values  $< 10^{-15}$ .

# Stochastic variability in promoter activity

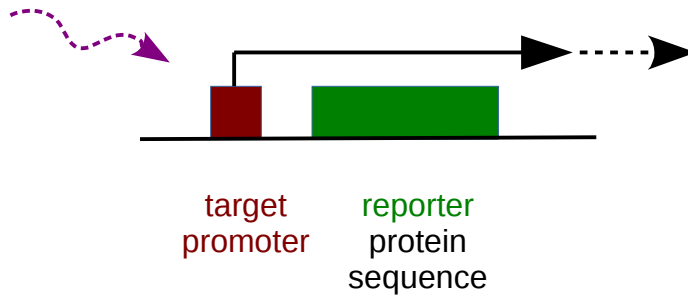


# Population snapshot data

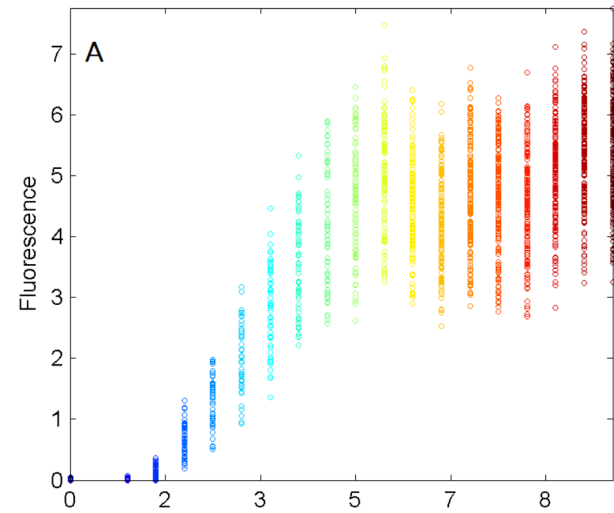
Gene of interest



Gene reporter system

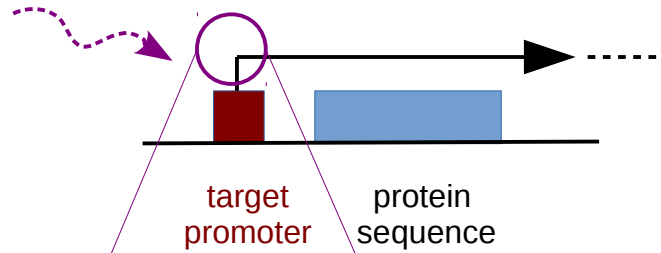


Fluorescence distribution in samples from a population of cells:

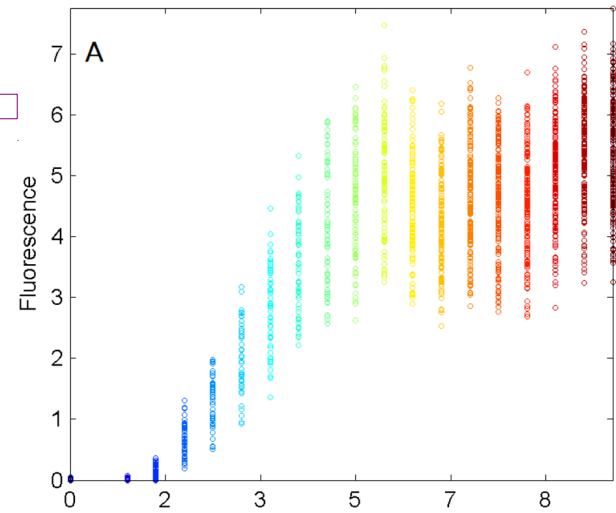
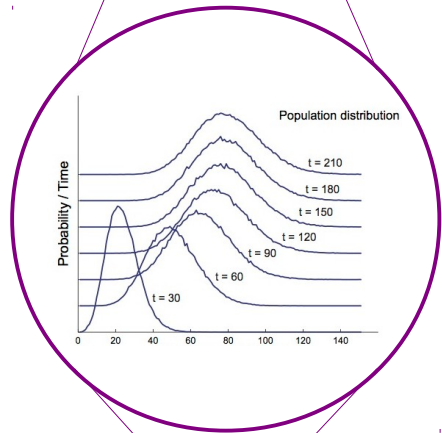
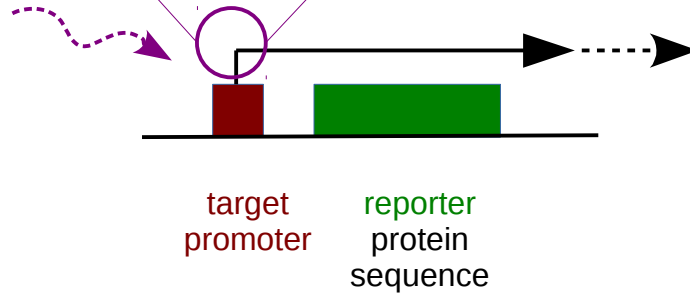


# Goal : Inference of promoter activity statistics

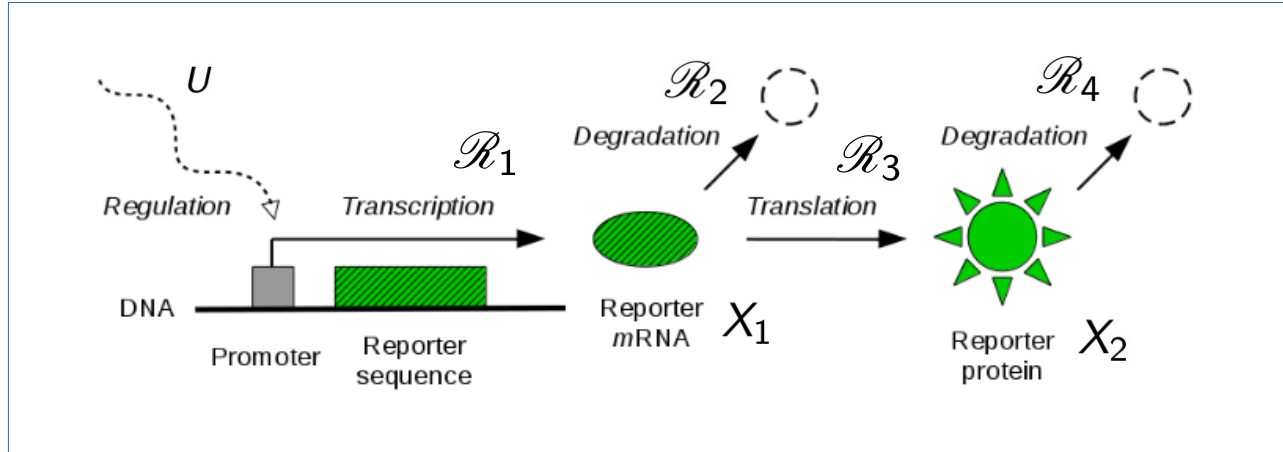
Gene of interest



Gene reporter system



# Random telegraph model

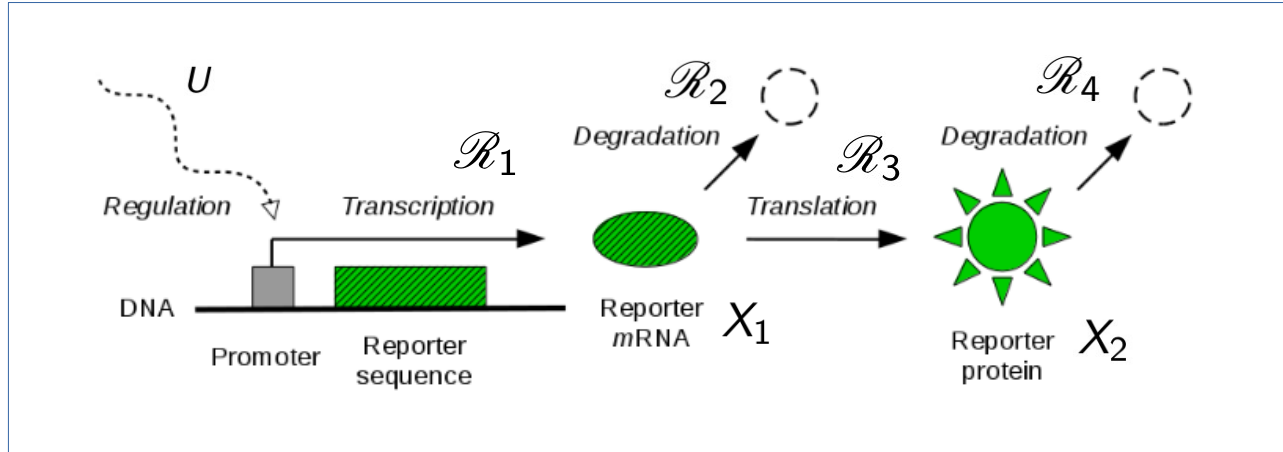


- Reactions :  $\mathcal{R}_1 : \emptyset \xrightarrow{k_M \cdot U} M$      $\mathcal{R}_2 : M \xrightarrow{d_M} \emptyset$   
 $\mathcal{R}_3 : M \xrightarrow{k_P} M + P$      $\mathcal{R}_4 : P \xrightarrow{d_P} \emptyset$

- Stoichiometry matrix and reaction rates :

$$S = \begin{bmatrix} 1 & -1 & 0 & 0 \\ 0 & 0 & 1 & -1 \end{bmatrix} \quad w(t) = \begin{bmatrix} k_M U(t) \\ d_M X_1(t) \\ k_P X_1(t) \\ d_P X_2(t) \end{bmatrix}$$

# Random telegraph model



- Reaction rates are affine in the state :

$$w(t) = WX(t) + F(t)$$

$$W = \begin{bmatrix} 0 & 0 \\ d_M & 0 \\ k_P & 0 \\ 0 & d_P \end{bmatrix} \quad F(t) = \begin{bmatrix} k_M U(t) \\ 0 \\ 0 \\ 0 \end{bmatrix}$$

# Moment Equations

- First- and second-order statistics of  $X$  :

$$\mu(t) = \mathbb{E}[X(t)], \quad \Sigma(t) = \text{Var}(X(t)), \quad \rho(z, t) = \text{Cov}(X(z), X(t))$$

- If  $U$  ( that is,  $F$  ) is a deterministic function :

$$d\mu(t)/dt = SW\mu(t) + SF(t)$$

$$d\Sigma(t)/dt = SW\Sigma(t) + \Sigma(t)W^T S^T + S\text{diag}(W\mu(t) + F(t))S^T$$

$$\partial\rho(z, t)/\partial z = SW\rho(z, t)$$

# Generalized Moment Equations

- Now assume  $F$  is a(ny) stochastic process :

$$\mu_F(t) = \mathbb{E}[F(t)], \quad \rho_F(z, t) = \text{Cov}(F(z), F(t)), \quad \xi_F(t) = \text{Cov}(X(0), F(t))$$

- Assuming *absence of feedback* from  $X$  to  $F$  :

$$\begin{aligned} d\mu(t)/dt &= SW\mu(t) + S\mu_F(t) \\ d\Sigma(t)/dt &= SW\Sigma(t) + \Sigma(t)W^T S^T + S\text{diag}(W\mu(t) + \mu_F(t))S^T \\ &\quad + V_{\xi_F}(t, t) + V_{\xi_F}^T(t, t) + V_{\rho_F}(t, t) + V_{\rho_F}(t, t)^T \\ \partial\rho(z, t)/\partial z &= SW\rho(z, t) + V_{\xi_F}(z, t) + V_{\rho_F}(z, t) \end{aligned}$$

$V_{\xi_F}$  linear (integral) functional of  $\xi_F$

$V_{\rho_F}$  linear (integral) functional of  $\rho_F$

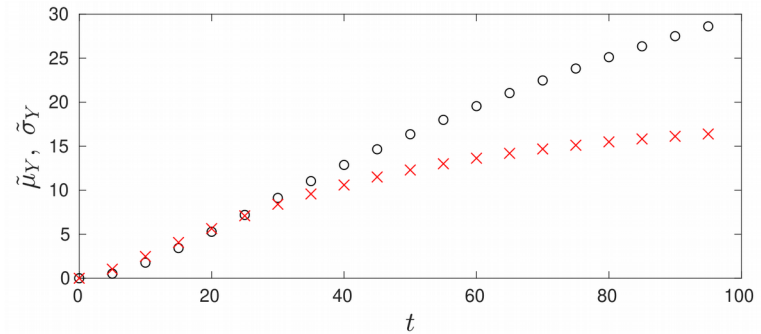


# Inference of promoter activity statistics

- Given empirical statistics (population snapshot data)

$$\tilde{\mu}_2(t_k) = \mu_2^*(t_k) + e_k^\mu$$

$$\tilde{\sigma}_2^2(t_k) = \Sigma_{2,2}^*(t_k) + e_k^\sigma$$



with  $k=1, \dots, K$  and i.i.d. approx Gaussian noise

- Estimate unknown mean and autocovariance function of  $U$
- Ill-posed linear inversion : Solve with (Tikhonov) regularization

# Special case : $U$ stationary, $X_0=0$

$$F(t) = \begin{bmatrix} k_M U(t) \\ 0 \\ 0 \\ 0 \end{bmatrix}$$

- Unknown stationary statistics :  $\bar{\mu}_U = \mu_U(\cdot)$ ,  $\bar{\rho}_U(\delta) = \rho_U(\cdot + \delta, \cdot)$
- Estimate constant mean by fitting mean data with the solution of

$$\frac{d}{dt}\mu(t) = SW\mu(t) + \begin{bmatrix} k_M \bar{\mu}_U \\ 0 \end{bmatrix}$$

- Estimate autocovariance as the solution of the convex optimization

$$\min_{\bar{\rho}_U \in \mathcal{C}} \sum_{k=1}^K \alpha_k^2 (\tilde{\sigma}_2^2(t_k) - \mathcal{L}(t_k | \bar{\rho}_U))^2 + \gamma \mathcal{Q}(\bar{\rho}_U)$$

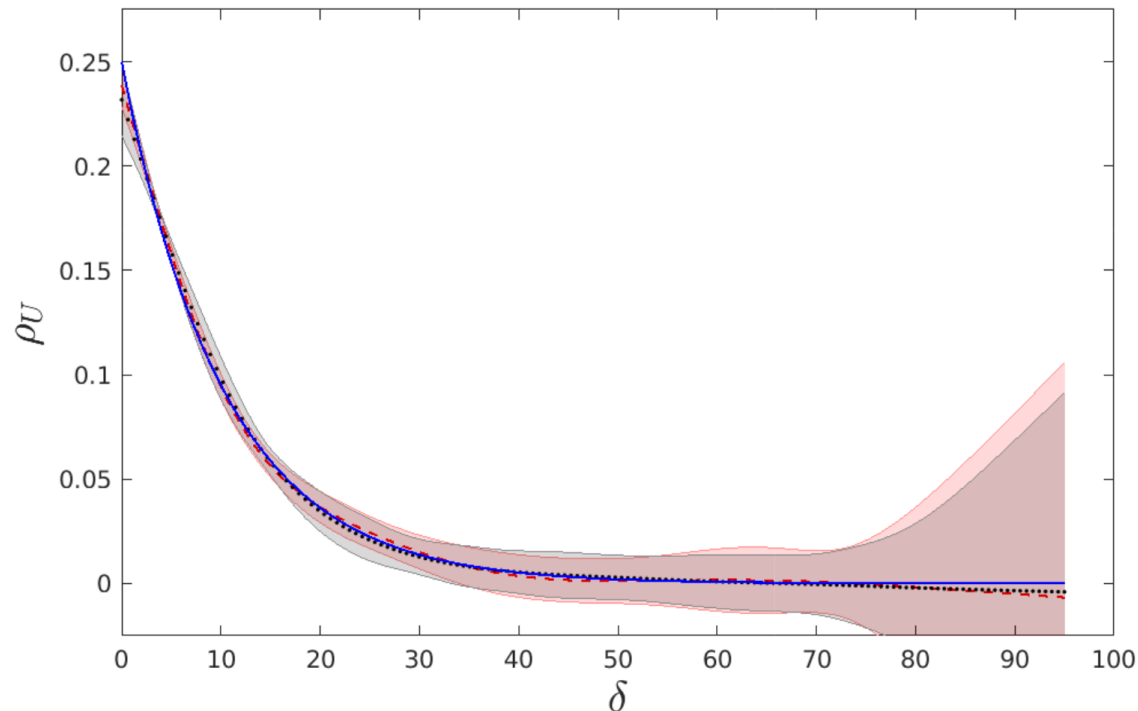
*PSD functions (convex cone)*      *Inverse of meas. error variance*      *Convex penalization of irregular solutions*

*Solution at  $t_k$  of  $d\Sigma(t)/dt = SW\Sigma(t) + \Sigma(t)W^T S^T + Q(t) + \Lambda(t|\bar{\rho}_U)$  with  $Q$  known function of the mean and  $\Lambda$  known functional*

- Implementation : Finite-dimensional LQP

# Results from numerical simulations

- Binary process  $U$  with stochastic switching rates
- Estimator mean  $\pm 2$  std for samples of  $10^5$  cells
- Automated vs. **fixed** choice of  $\gamma$  (vs. **true** autocovariance)



... Thanks !



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